## Environmental Monitoring and Cleaning<sup>1,2</sup>



## **Environmental Monitoring**

You should perform environmental monitoring each month with swab test samples collected from pre-PCR area surfaces such as centrifuges, pipettes, PCR racks, cold blocks, benchtops, and fridge and freezer doors. If environmental contamination is suspected, you should increase monitoring frequency.

Note: Do not swab in the post-PCR area. To reduce contamination and align with good molecular lab practices, do not bring samples from the post-PCR area into the pre-PCR area.

The recommended procedure for performing environmental monitoring swab tests is as follows:

- 1. Moisten a sterile swab applicator in a 1.5 mL microcentrifuge tube containing 250 μL nuclease-free water (one swab per surface area).
- 2. Wipe a test surface with the moistened swab back and forth a few times, rotating the swab as the sample is collected.
- 3. Put the swab back into the same microcentrifuge tube and agitate it in the water to release collected material.
- 4. Press the swab against the side of the tube while removing it to release excess water.
- 5. Discard the used swab.
- 6. Run the assay using an aliquot of the remaining eluant as if it is a purified sample.

## **Cleaning**

Laboratory cleaning should occur regularly. Additional cleaning may be necessary if swab test results indicate contamination. The most effective cleaning agent is a 10–15% bleach solution. Depending on the item, the recommended cleaning procedures are as follows:

For non-washable and non-removable items (i.e., pipettes and fridge/freezer doors):

- 1. Wipe the applicable surface with a 10%-15% bleach-dampened cleaning cloth.
- 2. Leave bleach on the surface for about 15 minutes.
- 3. Wipe the surface with a separate water-dampened cleaning cloth to remove bleach.
- 4. Wipe delicate items (such as pipettes) with 70% alcohol (if appropriate) for fast drying and leave others to air-dry.

For washable and removable items (i.e., PCR racks and centrifuge adaptors, not including pipettes):

- 1. Soak the applicable items in a 10-15% bleach solution for about 15 minutes.
- 2. Rinse treated items under running tap water to remove bleach.
- 3. Drain them on paper towels and leave to air dry, or wipe with 70% alcohol for fast drying if necessary.

Note: Alcohol drying is not required.

## REFERENCES

- 1. Aslanzadeh J. Preventing PCR amplification carryover contamination in a clinical laboratory. Ann Clin Lab Sci. 2004;34(4):389-96.
- 2. Nolan T, Huggett J, Sanchez E. Good practice guide for the application of quantitative PCR (qPCR). First Edition 2013. LGC (Internet). Cited 2021 May. Available from: http://www.gene-quantification.de/national-measurement-system-qpcr-guide.pdf.



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